



Soft Tissue Myoepitheliomas: A Discussion of Pathological Characteristics and Fusion Genes

Running Head: Pathology and Fusion Genes in Soft Tissue Myoepithelioma

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Abstract:

Owing to the diversity in cellular morphologies, nuclear atypia, proliferation patterns, and background matrices, rare soft tissue myoepitheliomas are often challenging to diagnose based solely on histopathological features. Benign tumors require differentiation from leiomyomas, neurogenic tumors, gastrointestinal stromal tumors, and solitary fibrous tumors. Due to histological similarities and overlaps, malignant myoepitheliomas should be differentiated from extraskeletal myxoid chondrosarcoma, proximal epithelioid sarcomas, extrarenal rhabdoid tumors, myoepithelioid tumor of the vulvar regions, and myoepithelioid tumor with chordoid features. Therefore, appropriate use of various markers, including myoepithelial cell markers and markers specific for other benign and malignant soft tissue tumors is essential.

To date, tumor-specific fusion genes have been identified in various soft tissue tumors, providing a useful complement to pathological diagnosis. Meanwhile, in soft tissue myoepithelioma, fusion genes composed of Ewing sarcoma breakpoint region 1 (EWSR1) or Fused in Sarcoma (FUS) gene with various partner genes and other fusion genes unrelated to EWSR1 or FUS have recently been identified. However, the relationship between these fusion genes and clinicopathological features has not been fully elucidated. Furthermore, therapeutic applications based on these findings have not yet been developed. Therefore, further research is needed to clarify them.

Keywords: Soft tissue myoepitheliomas; differential diagnosis; soft tissue tumors; fusion genes

Introduction:

Anatomically, myoepithelial cells are found in the ducts and acini of various glands, including the salivary, mammary, lacrimal, and sweat glands, and glands in the respiratory system. Myoepitheliomas usually arise in these organs and tissues [1-5], and rarely also occurring in soft tissues, bones, and viscera where myoepithelial cells are generally absent [6-9]. Such myoepitheliomas occur in a wide range of age groups, from infants to the elderly, and clinically exhibit a range of benign to low- to high-grade malignancies. Histologically, tumor cells exhibit oval to spindle shapes, mildly to severely atypical or pleomorphic nuclei, and a variety of growth patterns, including irregular fascicular, lobular, or glandular, often with diverse backgrounds such as fibrous, hyalinized, myxoid, or chondroid stroma. This heterogeneity and diversity in the histopathology [6-9] results in these histopathological similarities and overlaps with various other benign and malignant soft tissue tumors, making the pathological diagnosis extremely challenging. Therefore, to confirm the diagnosis of such soft tissue myoepithelioma, various myoepithelial markers, as well as markers specifically expressed in other soft tissue tumors require evaluation in addition to histological features.

In recent years, molecular and genetical studies, particularly of myoepithelioma arising in soft tissue and bone have revealed fusion genes related to Ewing sarcoma breakpoint region 1 (EWSR1) and Fused in Sarcoma (FUS), as well as several unrelated fusion genes [9-12]. However, the relationship between these fusion gene patterns and the clinicopathological features of rare soft myoepithelioma has not yet been fully elucidated. Additionally, therapeutic applications based on the fusion genes have not been developed. To resolve them, further accumulation of genetic data on this type of tumors is considered necessary.

Discussion

1. Pathological characteristics of soft tissue myoepitheliomas

Soft tissue myoepitheliomas display a wide clinical spectrum, ranging from benign to malignant. Histologically, similarities and overlaps with various benign and malignant soft tissue tumors are often observed, making a definitive diagnosis difficult. Benign tumors exhibit spindle-to-epithelioid cells and resemble benign soft tissue tumors, including leiomyomas, neurogenic tumors, gastrointestinal stromal tumors, and solitary fibrous tumors. In malignant cases, tumors exhibit more complex histopathological features, with tumor cells exhibiting diverse morphologies, including eosinophilic, amphophilic, or clear cytoplasm, moderate-to-severe nuclear atypia, pleomorphism, multinuclear structures, pathological mitotic figures, and even rhabdoid cells. Tumor cell nuclear loss of integrase interacting factor 1/switch/sucrose nonfermentable complex (SWI/SNF)-associated, matrix-associated, actin-dependent chromatin regulator subfamily b, member 1 (INI1/SMARCB1), and frequent positivity for marker of proliferation Ki-67 (Ki-67) and tumor protein p53 (p53) suggest a high-grade malignant myoepithelioma [13,14]. Additionally, the stroma of the background exhibits diverse appearances, including myxomatous, chondroid, fibrous, and hyalinized features. Regardless of benign or malignant form soft tissue myoepitheliomas are often positive tumor protein p63 (p63), S-100 protein (S-100), CD10, α -smooth muscle actin (SMA), caldesmon, calponin, and glial fibrillary acidic protein (GFAP). Use of these markers as well as various specific markers found in other benign and malignant soft tissue tumors complements and strengthens the pathological diagnosis of soft tissue myoepithelioma.

2. Differentiation between malignant soft tissue myoepitheliomas and soft tissue sarcomas

Complex features of malignant soft tissue myoepitheliomas suggest similarities and overlaps with various other malignant soft tissue tumors. Extraskeletal myxoid chondrosarcoma (EMC) usually shows mild to moderate nuclear atypia and is positive for vimentin, S100, epithelial membrane antigen (EMA), and neuroendocrine markers such as synaptophysin, cluster of



differentiation (CD) 56, and nuclear insulinoma-associated protein 1 (INSM1) [15, 16]. The rare, highly malignant cellular EMC consists of densely packed atypical epithelioid cells often with pleomorphic cells and rhabdoid cells often with nuclear loss of INI1/SMARCB1 [17]. Proximal epithelioid sarcoma (ES) is characterized by the cellular proliferation of highly atypical epithelial cells with numerous rhabdoid cells, usually without a myxoid background. The tumor cells were diffusely and strongly positive for vimentin, cytokeratin (CK), and CD34. In addition, tumor cell nuclear loss of INI1/SMARCB1 was observed in most cases [18]. Extrarenal rhabdoid tumor shows histological features very similar to those of proximal ES but are found in children, and the tumor cells are CD34-negative [19]. Recently, myoepithelioid tumor of the vulval region (METVR) [20] has been proposed as a low-grade malignant tumor that occurs in women in which epithelioid and spindle-shaped tumor cells with moderate nuclear atypia grow in a cord-like, reticular, or nest-like pattern against abundant myxoid stroma. This tumor occasionally harbors rhabdoid cells that show nuclear loss of INI1/SMARCB1. In addition, the tumor cells are estrogen receptor- and SMA-positive with a minority being CK-positive, but S-100, GFAP, and CD34-negative. Subsequently, myoepithelioid tumor with chordoid features [21] was described. This tumor shows histological features similar to those of METVR. However, unlike METVR, it occurs in both men and women with notochordal histological features with brachyury-positive tumor cells. Hence, to confirm the pathological diagnosis of malignant soft tissue myoepitheliomas, it is important to understand the histological and immunohistochemical characteristics of not only myoepitheliomas but also various other soft tissue tumors mentioned above.

3.Fusion genes in soft tissue mesotheliomas and salivary gland myoepitheliomas

In recent years, molecular and genetic studies have been conducted on various soft tissue tumors, and tumor-associated genetic abnormalities, particularly specific fusion genes, have been identified [22-24]. These findings may strengthen or complement the pathological diagnoses based on histological and immunohistochemical findings. In soft tissue myoepitheliomas, EWSR1 or FUS-related fusion genes have been detected individually with various partner genes [25-28]. In a notable study [9], EWSR1 and FUS fusion genes were examined in 66 cases of benign and malignant myoepitheliomas using fluorescent in situ hybridization (FISH), targeted RNA sequencing, and cytogenetics. EWSR1 was present in 54 cases (81.8%), and EUS was present in 12 cases (18.2%). The partner genes of EWSR1 were POU domain, class 5, transcript factor 1 (POU5F1) in 15 cases, Pre-B-cell leukemia transcription factor 3 (PBX3) in 10 cases, Pre-B-cell leukemia transcription factor 3 PBX1 in 6, Zinc finger protein 444 (ZNF444) in 3, Krueppel-like factor 15 (KLF15) in 2, and KLF17 in 1, but the partner gene was unknown in 17 cases. The partner genes of FUS were KLF17 in eight cases, POU5F1 in two cases, and unknown in 2. Of the 66 cases with EWSR1 and FUS rearrangements, 17 cases (25.8%) were malignant, consisting of EWSR1-POU5F1 (11/15, 73.3%), EWSR1-ZNF44 (3/3, 100%), and FUS-KLF17 (3/8, 37.5%). Additionally, EWSR1-POU5F1 malignant cases were mostly found in children and young adults, and histologically, they showed focal proliferation of clear epithelioid cells. EWSR1/FUS-PBX 1/3 of the cases were associated with benign with sclerosing spindle cell proliferation. Furthermore, various fusion genes unrelated to EWSR1 and FUS has been with the recent introduction of next generation sequencing (NGS). Fusion genes such as EWSR1-Vestigial-like family member 1 (VGLL1), Activating signal co-integrator 1 complex subunit 2 (ASCC2)-Nuclear factor erythroid 2 related factor1 (NBP2), Interferon regulatory factor 2 binding protein 2 (IRF2BP2)-Caudal type homeobox2 (CDX2), and Serum response factor (SRF)-E2F1 transcription factor 1 (E2F1), have been reported in soft tissue myoepitheliomas, ranging from benign to malignant [28-31]. However, the relationship between the fusion gene patterns and clinicopathological findings is not fully elucidated, requiring further studies including accumulation of the data. Myoepitheliomas arising from the salivary glands, sweat glands, glands in the respiratory tract, soft tissues, and other sites share common histopathological features. However, salivary gland myoepitheliomas usually harbor pleomorphic adenoma gene 1 (PLAG1)-related fusion genes [32-33]. These differ from the fusion genes found in soft tissue and cutaneous myoepitheliomas, including EWSR1 and FUS [9, 34], although some soft tissue myoepitheliomas harbor PLAG1-related fusion genes [35]. Furthermore, cutaneous syncytial myoepitheliomas harboring EWSR1-related fusion genes have been reported [34], and cutaneous myoepitheliomas with ductal differentiation have been associated with PLAG1-related fusion genes [36]. Indeed, there are overlaps in the fusion genes of salivary gland, skin, and soft tissue myoepitheliomas. Therefore, to clarify the relationship between clinicopathological findings and fusion gene patterns in soft tissue myoepitheliomas, detailed large-scale studies, including not only soft tissue

myoepitheliomas but also myoepitheliomas occurring in other tissues and organs are necessary. However, to date, no treatments for myoepitheliomas have been developed based on the detected fusion genes [37].

4.Treatment of soft tissue myoepithelioma based on fusion genes

Owing to its rarity of soft tissue myoepithelioma, there has been no consensus regarding chemotherapy or other systemic therapies for this tumor, so far. Meanwhile, a recent report showed that a combination of mitogen-protein kinase kinase (MEK) inhibitors and cobimetinib was effective for this tumor harboring Acylglycerol kinase (AGK)-B-Raf proto-oncogene, serine/threonine kinase (BRAF) fusion gene, based on the results of NGS [38]. Like this case, in the soft tissue myoepithelioma, it is expected to lead to the development of new treatments using NGS.

Conclusion

Rare soft tissue myoepitheliomas exhibit a wide spectrum of benignity and low- to high-grade malignancy with clinicopathological diversity, including biological behavior, cell morphology, nuclear atypia, proliferation patterns, and background matrices, often making pathological diagnosis challenging. Recent molecular and genetic studies in genetic abnormalities in soft tissue myoepitheliomas have detected fusion genes related to EWSR1 and FUS as well as a variety of unrelated fusion genes. However, the relationship between these fusion gene patterns and clinicopathological features has not been sufficiently elucidated. Therefore, large-scale studies are required to investigate these associations. Furthermore, later, it is necessary to aim to develop new treatments based on the findings from genetical research of this tumor.

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Conflict of Interest

The author declares no conflict of interest.

References:

1. [Telugu RB, Gaikwad P, Baitule A, Michael RC, Mani T, et al. \(2021\) Myoepithelial tumors of salivary gland: a clinicopathologic and immunohistochemical study of 15 patients with MIB-1 correlation. Head Neck Pathol. 15:479–490. 10.1007/s12105-020-01225-0](#)
2. [Huin M, Body G, Arbion F, Ouldamer L. \(2022\) Malignant breast myoepithelioma: a systematic review. Hum Reprod. 51:102481. 10.1016/j.jogoh.2022.102481. 10.1016/j.jogoh.2022.102481](#)
3. [Grossniklaus HE, Wojno TH, Wilson MW, Someren AO. \(1997\) Myoepithelioma of the lacrimal gland. Arch Ophthalmol. 115:1588–1590. 10.1001/archophth.1997.01100160758017](#)
4. [Hornick JL, Fletcher CD. \(2004\) Cutaneous myoepithelioma: a clinicopathologic and immunohistochemical study of 14 cases. Hum Pathol. 35:14–24. 10.1016/j.humpath.2003.08.016](#)
5. [Chand M, Mann JM, Sabayev V, Luo JJ, Cohen PR, et al. \(2011\) Endotracheal myoepithelioma. Chest. 140:242–244. 10.1378/chest.10-2976](#)
6. [Hornick JL, Fletcher CDM. \(2003\) Myoepithelial tumors of soft tissue: a clinicopathologic and immunohistochemical study of 101 cases with evaluation of prognostic parameters. Am J Surg Pathol. 27:1183–1196. 10.1097/0000478-200309000-00001](#)
7. [Verma A, Rekhi B. \(2017\) Myoepithelial tumor of soft tissue and bone: a current perspective. Histol Histopathol. 32:861–877. 10.14670/HH-11-879](#)
8. [Song W, Flucke U, Suurmeijer AIH. \(2017\) Myoepithelial tumors of bone. Surg Pathol Clin. 10:657–674. 10.1016/j.path.2017.04.010](#)
9. [Suurmeijer AIH, Dickson BC, Swanson D, Zhang L, Sung Y-S, et al. \(2020\) A morphologic and molecular reappraisal of myoepithelial tumors of soft tissue, bone, and viscera with EWSR1 and FUS gene rearrangements. Genes Chromosomes Cancer. 59:348–356. 10.1002/gcc.22835](#)
10. [Cyrtia J, Rosiene J, Bareja R, Kudman S, Zoughbi WA, et al. \(2022\)](#)



- [Whole-genome characterization of myoepithelial carcinomas of the soft tissue. Cold Spring Harb Mol Case Stud. 8:a006227. 10.1101/mcs.a006227](#)
11. [Patton A, Speeckaert A, Zeltman M, Cui X, Oghumu S, et al. \(2023\) A novel IRF2BP2::CDX22 Gene fusion in digital intravascular myoepithelioma of soft tissue: an enigma. Genes Chromosomes Cancer. 62:176–183. 10.1002/gcc.23108](#)
12. [Urbini M, Astolfi A, Indio V, Tarantino G, Serravalle S, et al. \(2017\) Identification of SRF- E2F1 fusion transcript in EWSR-negative myoepithelioma of the soft tissue. Oncotarget. 8:60036–60045. 10.18632/oncotarget.17958](#)
13. [Hollmann TJ, Hornick JL. \(2011\) INI1-deficient tumors: Diagnostic features and molecular genetics. Am J Surg Pathol. 35:e47–e63. 10.1097/PAS.0b013e31822b325b](#)
14. [Sonobe H, Omote R, Yukihiro K, Masumoto H, Yanai H, et al. \(2023\) Histopathological characterization of a rare case of soft tissue malignant myoepithelioma: a diagnostic challenge. HSOA J Clin Stud Med Case Rep. 2023; 10:17. 10.24966/CSMC-8801/1000178](#)
15. [Goh YW, Spagnolo DV, Platten M, Caterina P, Fisher C, et al. \(2001\) Extraskelatal myxoid chondrosarcoma: a light microscopic, immunohistochemical, ultrastructural and immunoultrastructural study indicating neuroendocrine differentiation. Histopathology. 39:514--524. 10.1046/j.1365-2559.2001.01277.x](#)
16. [Yoshida A, Makise N, Wakai S, Kawai A, Hiraoka N. \(2018\) INSM1 expression and its diagnostic significance in extraskelatal myxoid chondrosarcoma. Mod Pathol. 2018; 31:744–752. 10.1038/modpathol.2017.189](#)
17. [Flucke U, Tops BB, Verdiijk MA, van Cleef PIH, van Zwam PH, et. al. \(2012\) NR4A3 rearrangement reliably distinguishes between the clinicopathologically overlapping entities myoepithelial carcinoma of soft tissue and cellular extraskelatal myxoid chondrosarcoma. Virchows Arch. 460:621–628. 10.1007/s00428-012-1240-0](#)
18. [Hornick JL, Dal Cin P, Fletcher CD. \(2009\) Loss of INI1 expression is characteristic of both conventional and proximal type epithelioid sarcoma. Am J Surg Pathol. 33:542–550. 10.1097/PAS.0b013e3181882c54](#)
19. [Fanburg-Smith JC, Hengge M, Hengge UR, et al Smith Jr JS, Miettinen M. \(1998\) Extrarenal rhabdoid tumors of soft tissue: a clinicopathologic and immunohistochemical study of 18 cases. Ann Diagn Pathol. 1998; 2:351–362. 10.1016/s1092-9134\(98\)80038-5](#)
20. [Yoshida A, Yoshida H, Yoshida M, Mori T, Kobayashi E, et. al. \(2015\) Myoepithelioma-like tumors of the vulvar region: a distinctive group of SMARCB1-deficient neoplasms. Am J Surg Pathol. 2015; 39:1102–1113. 10.1097/PAS.0000000000000466](#)
21. [Kinoshita I, Kohashi K, Yamamoto H, Yamada Y, Inoue T, et. al. \(2021\) Myxoepithelioid tumour with chordoid features: a clinicopathological, immunohistochemical and genetic study of 14 cases of SMARCB1/INI1-deficient soft-tissue neoplasm. Histopathology. 79:629–641. 10.1111/his.14393](#)
22. [Fisher C. \(2014\) The diversity of soft tissue tumours with EWSR1 gene rearrangements: a review. Histopathology. 64:134–150. 10.1111/his.12269](#)
23. [23. Flucke U, van Noesel MM, Siozopoulou V, Creytens D, Tops BBI, et. al. \(2021\) EWSR1-The most common rearranged gene in soft tissue lesions, which also occurs in different bone lesions: an updated review. Diagnostics \(Basel\). 11:1093. 10.3390/diagnostics11061093](#)
24. [Antonescu CR, Zhang L, Chang N-E, Pawel BR, Travis W, et al. \(2010\) EWSR1-POU5F1 fusion in soft tissue myoepithelial tumors. A molecular analysis of sixty-six cases, including soft tissue, bone, and visceral lesions, showing common involvement of the EWSR1 gene. Genes Chromosomes Cancer. 49:1114–1124. 10.1002/gcc.20819](#)
25. [Huang SC, Chen HW, Zhang L, Sung Y-S, Agaram NP, et al. \(2015\) Novel FUS-KLF17 and EWSR1-KLF17 fusions in myoepithelial tumors. Genes Chromosomes Cancer. 54:267–275. 10.1002/gcc.22240](#)
26. [Leduc C, Zhang L, Öz B, Luo J, Fukuoka J, et. al. \(2016\) Thoracic myoepithelial tumors: a pathologic and molecular study of 8 cases with review of the literature. Am J Surg Pathol. 40:212–223. 10.1097/PAS.0000000000000560](#)
27. [Komatsu M, Kawamoto T, Kanzawa M, Kawakami Y, Hara H, et. al. \(2020\) A novel EWSR1-VGLL1 gene fusion in a soft tissue malignant myoepithelial tumor. Genes Chromosomes Cancer. 59:249–254. 10.1002/gcc.22823](#)
28. [Leckey BD Jr, John I, Reyes-Múgica M, Naous R. \(2021\) EWSR1-ATF1 Fusion in a myoepithelial carcinoma of soft tissue with small round cell morphology: a potential diagnostic pitfall. Pediatr Dev Pathol. 24:258–263. 0.1177/1093526621998869](#)
29. [Cyrtá I, Rosiene J, Bareja R, Kudman S, Zoughbi WA, et. all. \(2022\) Whole-genome characterization of myoepithelial carcinomas of the soft tissue. Cold Spring Harb Mol Case Stud. 8:a006227. 10.1101/mcs.a006227](#)
30. [Patton A, Speeckaert A, Zeltman M, Cui X, Oghumu S, et al. \(2023\) A novel IRF2BP2::CDX22Gene fusion in digital intravascular myoepithelioma of soft tissue: an enigma. Genes Chromosomes Cancer. 62:176–183. 10.1002/gcc.23108](#)
31. [Urbini M, Astolfi A, Indio V, Tarantino G, Serravalle S, et. al. \(2017\) Identification of SRF- E2F1 fusion transcript in EWSR-negative myoepithelioma of the soft tissue. Oncotarget. 8:60036–60045. 10.18632/oncotarget.17958](#)
32. [Rupp NJ, Höller S, Brada M, Vital D, Morand GB, et. al. \(2022\) Expanding the clinicopathological spectrum of TGFBR3-PLAG1 rearranged salivary gland neoplasms with myoepithelial differentiation including evidence of high-grade transformation. Genes Chromosomes Cancer. 61:94–104. 0.1002/gcc.23009](#)
33. [Skálová A, Agaimy A, Vanecek T, Banekova M, Laco J, et. al. \(2021\) Molecular profiling of clear cell myoepithelial carcinoma of salivary glands with EWSR1 rearrangement identifies frequent PLAG1 gene fusions but no EWSR1 fusion transcripts. Am J Surg Pathol. 45:1–13. 10.1097/PAS.0000000000001591](#)
34. [Jo VY, Antonescu CR, Dickson BC, Swanson D, Zhang L, et. al. \(2020\) Cutaneous syncytial myoepithelioma is characterized by recurrent EWSR1- PBX3 fusions. Am J Surg Pathol. 43:1349–1354. 10.1097/PAS.0000000000001286](#)
35. [Segawa K, Sugita S, Aoyama T, Takenami T, Asanuma H, et. al. \(2020\) Myoepithelioma of soft tissue and bone, and myoepithelioma-like tumors of the vulvar region: Clinicopathological study of 15 cases by PLAG1 immunohistochemistry. Pathol Int. 70:965--974. 10.1111/pin.13017](#)
36. [Antonescu CR, Zhang L, Shao SY, Mosquera J-M, Weinreb I, et. al. \(2013\) Frequent PLAG1 gene rearrangements in skin and soft tissue myoepithelioma with ductal differentiation. Genes Chromosomes Cancer. 52:675–82. 10.1002/gcc.22063](#)
37. [Domen A, Paesschen CV, Zwaenepoel K, Lambin S, Pauwels P, et. al. \(2022\) Excellent Response to MEK Inhibition in an AGK-BRAF Gene Fusion Driven Carcinoma: Case Report and Literature Review. Anticancer Res. 42:373--379. 10.21873/anticancer.15495](#)
38. [38. Domen A, Paesschen CV, Zwaenepoel K, Lambin S, et al. \(2022\). Excellent Response to MEK Inhibition in an AGK-BRAF Gene Fusion Driven Carcinoma: Case Report and Literature Review. Anticancer Res. 42:373-379. 10.21873/anticancer.15495](#)



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